

Using IFCB to monitor filamentous cyanobacteria in the Baltic Sea

Automated data pipeline and CNN based classification system

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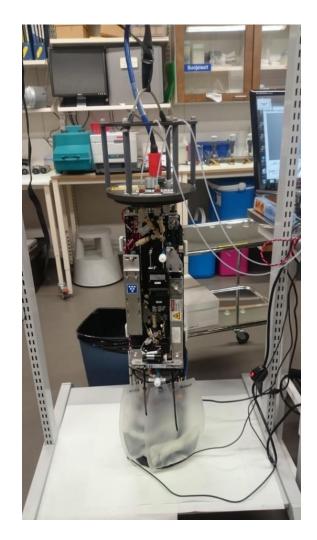
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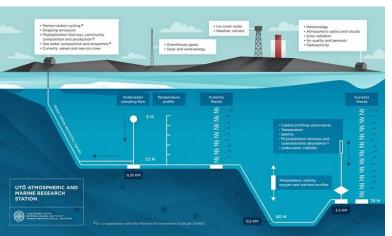
Imaging FlowCytobot - IFCB

- Imaging flow cytometer
- Takes images of phytoplankton cells and colonies inside the size range of 10-150 μm
- Can be operated remotely mounted on a flow through system at a research station (like in Utö), ship of opportunity, put to measure directly to the sea or used in a laboratory environment etc..
- Takes a sample of 5ml with approx. 20 min interval
- The camera is triggered by <u>chlorophyll-a</u> or scatter
- Even as many as ~30 000 high resolution images / hour
- 150 μ m mesh in IFCB inlet to prevent it from clocking
- Data analyzed with image recognition algorithm

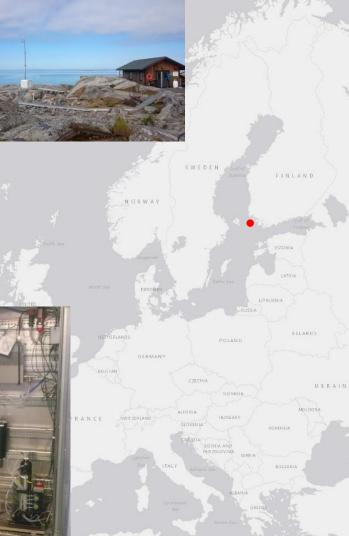


UTÖ Marine Research Station joint with Finnish Meteorological Institute

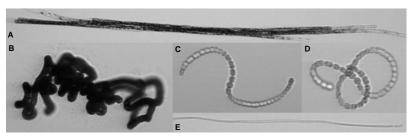
- Underwater pump with inlet at 5 m depth, 250 m offshore
- Water distributed to different sensors inside the station cabin
- Represents pelagial community of a mixed surface layer in brackish environment (salinity ~ 6 psu)
- Multiple parallel measurements from sea to atmosphere
- Continuous imaging flow cytometer observations from multiple years
- Light microscopy samples of cyanobacteria bloom in 2018







Cyanobacteria blooms in the Baltic Sea



Common filamentous species

- Aphanizomenon flosaquae (A) .
- *Dolichospermum* spp. (C,D) .
- Nodularia spumigena (B) .
- Oscillatoriales (E) .

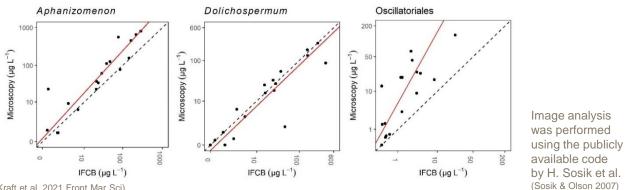


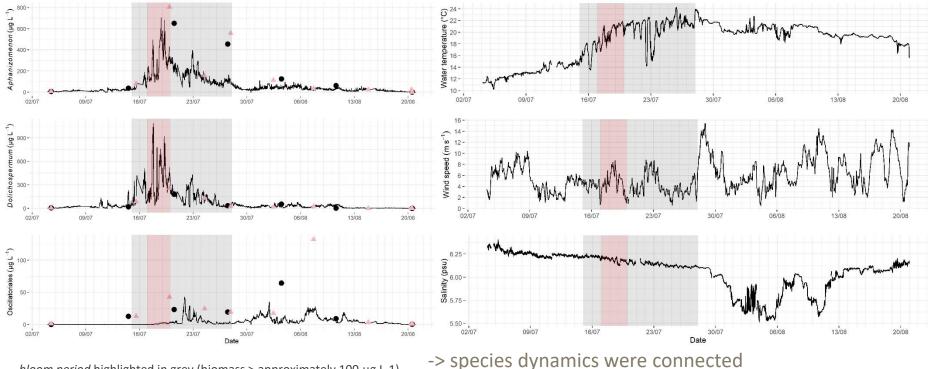






Photo by Lauri Laakso

High frequency phytoplankton observations connected with changes in the environment can help to understand mechanisms of the blooms

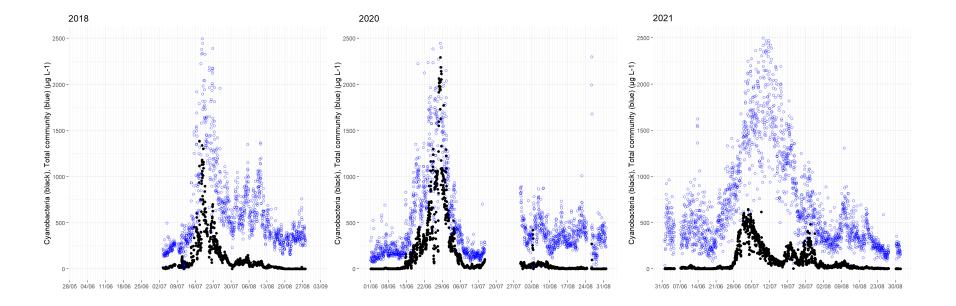


bloom period highlighted in grey (biomass > approximately 100 μg L-1) bloom peak period highlighted in red (the days with the highest peaks) light microscopy counts •flow-through ▲ pump inlet

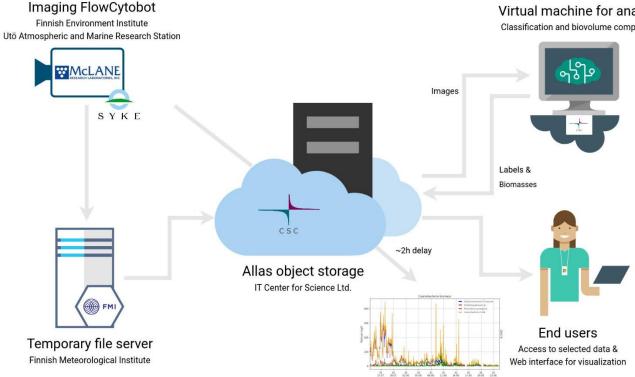
to changes in the water mass

(Kraft et al. 2021 Front Mar Sci)

Cyanobacteria blooms in 2018, 2020, 2021



Near-real time classification via **Convolutional Neural Networks**



https://github.com/veot/syke-pic, Kraft et al. 2022 Front Mar Ski

Virtual machine for analysis Classification and biovolume computation

- Training data set •
- http://doi.org/10.23728/b2share.abf . 913e5a6ad47e6baa273ae0ed6617a
- Evaluation data set .
- http://doi.org/10.23728/b2share.7c . 273b6f409c47e98a868d6517be3ae3
- Visualization still under development •
- https://plankton.live/ .
- https://ifcb.plankton.live/timeline?da . taset=uto
- https://swell.fmi.fi/hab-info/ •

Classifier performance

- Kraft et al. 2022 Front Mar Sci
- Weighted F1-score of test data of our labeled image data set 0.95
- Weighted F1-score of our evaluation data (59 natural samples annotated entirely) 0.83
- Class-specific thresholds are used for filtering out the unidentifiable images, determined with our test set complemented with unidentifiable images (initial situation -> to be finetuned as data accumulates)
- ND in the Evaluation Data means "Not Determined", the metrics were not calculated for classes with <10 images

Table contains results of evaluation data for F1 > 0.7 Gymnodi

	Training	Validation	Test				2021 data			
Class / taxonomic group	N	Thre	N	Pr	Re	F1	N	Pr	Re	F1
Oscillatoriales	2664	0.31	888	0,99	1,00	0,99	3893	0,98	0,98	0,98
Monoraphidium contortum	196	0.69	66	0,98	0,98	0,98	439	0,99	0,96	0,97
Skeletonema marinoi	2477	0.46	825	1,00	0,99	0,99	7402	0,99	0,94	0,97
Heterocapsa triquetra	1966	0.39	655	0,98	0,97	0,97	2267	0,92	0,95	0,94
Cryptophyceae / Teleaulax sp.	4098	0.53	1366	0,96	0,97	0,96	16952	0,97	0,90	0,93
Aphanizomenon flosaquae	4193	0.24	1398	0,97	1,00	0,98	1849	0,87	0,98	0,92
Peridiniella catenata chain	116	0.7	38	0,97	1,00	0,99	89	0,99	0,87	0,92
Dolichospermum sp. / Anabaenopsis sp.	7368	0.38	2456	0,98	0,99	0,98	790	0,88	0,96	0,92
Pauliella taeniata	71	0.62	24	1,00	0,96	0,98	56	0,96	0,86	0,91
Oocystis sp.	505	0.5	169	0,88	0,93	0,90	161	0,91	0,89	0,90
Mesodinium rubrum	679	0.44	227	0,96	0,95	0,96	560	0,92	0,86	0,89
Melosira arctica	26	0.3	8	0,73	1,00	0,84	58	0,85	0,91	0,88
Dolichospermum sp. / Anabaenopsis sp. coiled	1502	0.41	501	0,93	0,96	0,95	70	0,74	0,99	0,85
Eutreptiella sp.	1348	0.43	450	0,95	0,94	0,94	1678	0,90	0,76	0,83
Licmophora sp.	44	0.43	15	1,00	0,80	0,89	78	0,90	0,77	0,83
Nodularia spumigena	101	0.32	34	0,80	0,94	0,86	62	0,80	0,85	0,83
Heterocapsa rotundata	368	0.56	123	0,84	0,90	0,87	2609	0,89	0,70	0,78
Ceratoneis closterium	27	0.41	9	1,00	1,00	1,00	75	0,68	0,91	0,78
Peridiniella catenata single	539	0.52	180	0,89	0,97	0,93	222	0,75	0,81	0,78
Pennales thick	126	0.37	42	0,93	0,88	0,90	1088	0,72	0,85	0,78
Thalassiosira levanderi	1522	0.63	508	0,95	0,95	0,95	2008	0,87	0,68	0,77
Chaetoceros sp. chain	829	0.51	277	0,93	0,95	0,94	693	0,76	0,77	0,76
Centrales	288	0.51	96	0,98	0,89	0,93	92	0,77	0,68	0,72
Dinophysis acuminata	130	0.68	44	0,98	0,91	0,94	17	0,79	0,65	0,71
Pennales thin	469	0.29	156	0,96	0,99	0,97	334	0,61	0,84	0,71
Cyclotella choctawhatcheeana	61	0.47	21	0,89	0,81	0,85	199	0,92	0,57	0,71
7 Gymnodiniales	41	0.29	14	0,92	0,86	0,89	38	0,78	0,64	0,70

Group specific confusion of our evaluation data

Actual label	Cyanophyceae-	0.94	0	0	0	0	0	0	0	0.06
	Cryptophyceae-		0.89	0	0	0	0	0	0	0.11
	Euglenophyceae -		0.02	0.76	0.01	0	0	0	0	0.22
	Dinophyceae -		0		0.75	0	0		0	0.24
	Bacillariophyceae-	0.01	0	0	0	0.86	0	0	0	0.13
	Chrysophyceae-		0	0	0	0	0.51	0	0	0.49
	Chlorophyta-		0	0	0	0	0	0.34	0	0.65
	Ciliophora-		0	0	0.02	0.03	0	0	0.76	0.19
		Cyanophyceae -	Cryptophyceae -	Euglenophyceae -	Dinophyceae -	apoilariophyceae -	lade	Chlorophyta -	Ciliophora -	Unclassified -

What can we identify? ~50 categories, ~60000+ annotated images

Taxons / groups so far identified with IFCB from the Baltic Sea (work ongoing)

Cyanophyceae

Aphanothece paralleliformis Chroococcales Chroococcus spp. Merismopedia spp. Snowella spp. / Woronichinia spp. Oscillatoriales Aphanizomenon flosaquae Aphanizomenon spp. Dolichospermum spp. /Anabaenopsis spp. Nodularia spumiqena

Cryptophyceae

Cryptomonadales *Teleaulax* spp.

Dinophyceae

Prorocentrum cordatum Dinophyceae (under & over 20µm) Dinophysis acuminata Dinophysis norvegica Gymnodiniales Gymnodinium spp. Gymnodinium like cells Gyrodinium spp. Heterocapsa rotundata Heterocapsa triquetra Amylax triacantha Gonyaulax verior Peridiniella catenata Protoceratium reticulatum

Chrysophyceae

Dinobryon balticum Uroglenopsis spp. Apedinella radians Pseudopedinella spp.

Diatomophyceae

Chaetoceros spp. Chaetoceros danicus Chaetoceros similis Chaetoceros subtilis Chaetoceros throndsenii Coscinodiscus granii Cylindrotheca closterium Cvclotella choctawhatcheeana Melosira arctica Skeletonema marinoi Thalassiosira baltica Thalassiosira levanderi Diatoma tenuis Licmophora spp. Nitzschia paleacea Pauliella taeniata

Euglenophyceae Eutreptiella spp.

Chlorophyta Cymbomonas tetramitiformis Pyramimonas spp. Chlorococcales Scenedesmus spp. / Acutodesmus spp. / Desmodesmus spp. Monoraphidium contortum Binuclearia lauterbornii Oocystis spp.

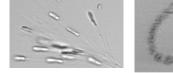
Katablepharis remigera Flagellates Nanoplankton

Ciliates Strombidium spp. Mesodinium rubrum

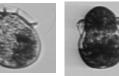
Additional categories include eg. akinetes heterocytes resting stages cysts

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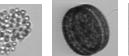


















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References and sources

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- Kraft et al. 2022. Towards operational phytoplankton recognition with automated high-throughput imaging, near real-time data processing, and convolutional neural networks. Frontiers in Marine Science, 9.
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- Olson & Sosik 2007. A submersible imaging-in-flow instrument to analyze nano-and microplankton: Imaging FlowCytobot. Limnology and Oceanography: Methods, 5(6), 195-203.
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- <u>https://www.finmari-infrastructure.fi/field-stations/uto-fmi/</u>
- <u>https://mclanelabs.com/imaging-flowcytobot/</u>